

In Depth Assessment of Off-target Editing by CRISPR/Cas9 in VOR33, an Engineered Hematopoietic Stem Cell Transplant for the Treatment of Acute Myeloid Leukemia

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INTRODUCTION

- VOR33 is an engineered allogeneic hematopoietic stem cell (HSC) transplant for treatment of acute myeloid leukemia (AML) in which the CD33 surface antigen is removed by CRISPR/Cas9 gene editing (Figure 1).¹
- This removal enables post-engraftment immunotherapeutic targeting of leukemic cells that display CD33 while sparing the CD33 gene-edited graft (Figure 2).^{2,4}
- To ensure safety of gene-edited CD33+ hematopoietic stem and progenitor cells (HSPCs) and engrafted progeny, a well-defined analyses of unintended and off-target editing is necessary. However, paradigms for off-target analyses of gene-edited ex vivo therapies are not well established.

Figure 1. VOR33 Engineering

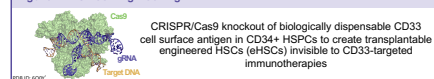
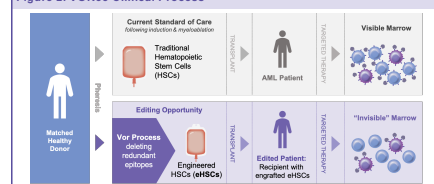


Figure 2. VOR33 Clinical Process



OBJECTIVE

- To enable rigorous assessment of unintended and off-target editing events by CRISPR/Cas9 in VOR33 using an ensemble of sensitive genomic assays and approaches.

OFF-TARGET STRATEGY

Potential Off-Target Concerns	Analytic Approach
Unintended on-target structural variation (SV)	Long-range PCR and long-read DNA sequencing (Figure 3)
Off-target sites with high homology to CD33 on-target site	<i>In silico</i> prediction of possible genomic sites ≤ 5 mismatches (Figure 4)
Off-target sites with moderate/poor homology to CD33 on-target site	Unbiased identification by GUIDE-seq (Figure 5)
Quantifying off-target indel frequencies	Hybrid capture-based next generation sequencing (NGS) (Figure 6)
Gross genomic instability	G-banded karyotyping (Figure 7)

Figure 3. Unintended On-Target SV Detection: Long-Range PCR and Long-Read Sequencing of On-Target CD33 site

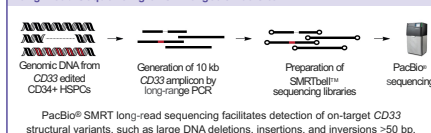


Figure 4. *In Silico* Off-Target Prediction

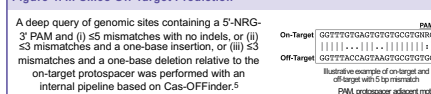


Figure 5. Unbiased Identification of Off-Target Sites: GUIDE-seq®

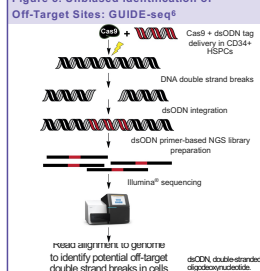
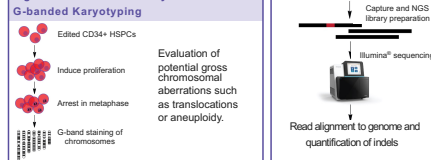


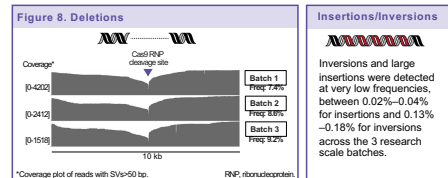
Figure 7. Genomic Stability Detection: G-banded Karyotyping



RESULTS

- Analysis of on-target SVs by long-read sequencing revealed total frequencies of 7%–9% across 3 VOR33 research-scale batches. Large deletions account for the majority of SVs. Large on-target inversions and insertions were detected at very low frequencies, similar to those previously reported for CRISPR/Cas9 (Figure 8).⁶ Fine mapping of the SVs suggests no perceivable impact on the safety or efficacy of VOR33, as the primary mechanism of action (MOA) by CD33 disruption is preserved.
- By conducting GUIDE-seq analysis on 4 research-scale batches, a total of 29 sites were identified, with 10 showing high homology to the on-target site (≤ 5 mismatch/gaps) (Table 1). The remaining 19 sites had moderate/poor homology (≥ 7 mismatch/gaps).
- In 4 research scale VOR33 batches, indel frequencies were assessed by hybrid capture-based NGS at >2300 *in silico* predicted sites (Table 2). In 7 VOR33 batches manufactured at clinical scale, indel frequencies were assessed by NGS at 2369 *in silico* and GUIDE-seq identified sites (reads ≥ 500) (Table 3). Across batches, no significant and reproducible off-target sites were observed.
- Lastly, karyotyping revealed no detectable abnormalities across multiple research and clinical scale batches, indicating that VOR33 displays preserved genomic stability (Figure 9, Figure 10).

Unintended On-Target Structural Variant Characterization



Unbiased Prediction of Off-Target Sites

Table 1. GUIDE-seq Predicted Off-Target Sites From 4 Research Scale Batches With 2 dODN Concentrations (8 Independent Samples)

Mismatches/gaps	Counts
3	8
4	1
5	1
6	2
7 and above	19

CONCLUSIONS

- The VOR33 engineering process is robust and reproducible, with no discernable differences in off-target frequencies or patterns in multiple independent cell batches generated with various guide RNA lots, manufacturing scales, and delivery methods.
- An expansive appraisal of off-target editing across multiple VOR33 products was achieved through long-read sequencing, GUIDE-seq, quantification of indel frequencies at >2300 genomic sites, and karyotyping.
- This assessment of off-target editing establishes a rigorous and clinically translatable safety framework to evaluate genotoxicity in CD34+ HSPC-based cell therapies for the treatment of AML.

References
 1. Zhu X, et al. *Nat Struct Mol Biol*. 2019;26(8):679-685. 2. Borot F, et al. *Proc Natl Acad Sci U S A*. 2019;116(24):11978-11987. 3. Humbert O, et al. *Sci Transl Med*. 2019;11(503):eaaw3768. 4. Kim MY, et al. *Cell*. 2018;173(6):1439-1453.e19. 5. Bao S, et al. *Bioinformatics*. 2014;30(10):1473-1475. 6. Tsai SQ, et al. *Nat Biotechnol*. 2015;33:187-197. 7. Grise A, et al. *Nat Biotechnol*. 2020;27:182-189. 8. Kowalski M, et al. *Nat Biotechnol*. 2018;36(8):765-771. 9. Chaudhuri HG, et al. *CRISPR J*. 2020;3(6):440-453. Content illustrations created with BioRender.com.

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 See the companion presentation: Pre-clinical evaluation, including genomic off-target analysis, of VOR33: a clinically-ready CRISPR/Cas9 engineered hematopoietic stem cell transplant for the treatment of acute myeloid leukemia (CR36).

